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Ramanou AA et al, The Experiment, 2014., Vol. 27(1), 1844-1851

ANTIBACTERIAL POTENCIES OF TEN IVORIAN MEDICINAL PLANTS AGAINST MULTI DRUG RESISTANT STRAINS OF SALMONELLA

ABSTRACT

Aim: The purpose of this study was to evaluate the antisalmonella activity of the aqueous and 70% ethanol extract of ten (10) Ivorian plants used in traditional medicine to cure infectious diseases against two (2) clinical multi-drug resistant strains of Salmonella typhiand Salmonella typhimurium.

Methods: The antimicrobial parameters (Minimal inhibitory Concentration MIC and Minimal Bactericidal Concentration MBC) were determined by the broth dilutionmethod and the diameters of inhibition zone were determined by agar disc diffusion method.

Results: The aqueous extract of Thonningia sanguineand 70% ethanol extract of Abrusprecatorius had the same and the best MIC and MBC values respectively 5mg/ml and 10mg/ml for the multi drug resistant strains of Salmonella tested. Moreover, the inhibition diameters indicate that the aqueous extract of Thonningia sanguine showed the best significant activity against the multi-drug resistant strain of Salmonella.typhimurium ($11\pm 0, 57$).

Conclusion: The aqueous extract of Thonningiasanguinea and 70% ethanol extract of Abrusprecatorius can provide an alternative therapy for the treatment of salmonellosis generated by multi drug resistant strains.

Key words: Minimal Inhibitory Concentration, Minimal Bactericidal Concentration, antimicrobial activity, Thonningiasanguinea, Abrusprecatorius

INTRODUCTION

Salmonella Spp is a primary cause of food poisoning worldwide. The center for disease control and prevention estimated that approximately 1.4millions cases of salmonellosis were annually reported in the United States¹. It is also a public health problem in developing countries. SalmonellaentericaserovarTyphi causes approximatively 10 million cases oftyphoid fevereach year mostly in developing countries^{2, 3}. In developing countries, enteric fever (typhoid) is more severe due to poor hygiene, indiscriminate use of antibiotics, and a rapid rise in mutidrugresistance. In Côte d'Ivoire (West Africa),the typhoid fever and the over forms of salmonellosis became these last years a problem of public health taking into account the degradation of the conditions of healthiness in relation to the civil war occurred from 2002 to 2011. In recent years there has been a rapid rise in multidrug resistance by Salmonella typhi all over the world^{4, 5, 6}. The world health organisation estimated an annual rate of 12, 6 million typhoid fever infections with nearly 600000 deaths every year⁷. Resistance to the first line drugs, chloramphenicol, ciprofloxacin and amoxicillin, in the course of salmonellosis management has been reported⁸. The pathogenic role of salmonella infection in the development of human diseases and the impact of resistance on the clinical outcome stimulated the search for newer treatments and natural products whichcould provide alternative therapies against salmonellosis.

Previous results revealed the antibacterial activities, with MIC varying from 5 mg/mL to 40 mg/mL, of aqueous and ethanolic extracts of ten Ivorian medicinal plants against two sensitive strains of Salmonella typhi and Salmonella typhimurium⁹. At the best of our knowledge, none study had been done to assess the sensibility of MDR strains of salmonella with the same plants except the aqueous extract of Thonningiasanguinea on Salmonellatyphi DT 104¹⁰.

With regard to foregoing considerations, the present study was undertaken to evaluate the antibacterial activity of the ten plants against two clinical MDR strains of salmonella. The minimal inhibitory concentrations (MIC) and the minimal bactericidal concentrations (MBC) were determined for the aqueous and 70% ethanol extracts of each plant.

RESEARCH ARTICLE



Ramanou AA et al, The Experiment, 2014., Vol. 27(1), 1844-1851

MATERIAL AND METHODS Collection of plant material

All plants were collected from the wild in different area of Côte d'Ivoire and were identified and authenticated by PrAkéAssi of the department of Botany, university of Cocody-Abidjan. Vouchers specimens were deposited in the herbarium of "National Floristic Center" of Abidjan. The plant species, parts used, local name, voucher specimen numbers and the traditional uses of the plants are listed in Table 1.

Bacterial strains

Bacteria (clinical strains) for testing purposes were kindly provided by the National Laboratory of Public Health of Côte d'Ivoire. Salmonella typhi resistant to chloramphenicol and cefotaxim Salmonella typhimurium resistant to cefotaxim and ceftriaxon They were sub cultured on nutrient agar for 15 days and maintained on nutrient agar slants at 4°C. Fresh inoculums were taken for test.

Preparation of aqueous extracts

The plant extracts were prepared using the method of Guédé-Guinaet al.¹¹. The freshly collected flowers, leaves and seeds of the plants were air dried at room temperature for 7 days and powdered. Briefly 20g of powder were soaked in 500mL distilled water for 24h with constant stirring. The suspension was further filtered through clean sterile muslin cloth and watmann N°1 filter paper. The filtrate was concentrated in vacuum using a rotary evaporator to obtain the aqueous extract.

Preparation of 70% ethanol extracts

The plant extracts were prepared using the method of Zirihiet al.¹². Briefly 20g of powder were soaked in 300ml 70% ethanol [ethanol/water (70/30, V/V)]. The mixture was stirred during 24 hours and then filtered through a clean sterile muslin cloth. The filtrate was decanted for 24h. The hydroalcoolicsolutionwas filtered through watmann N°1 filter paper and concentrated in vacuum at 40°C to obtain the 70% ethanolic extract.

Preparation of different fractions

Twenty (20)g of aqueous extract of Thonningiasangineaand 70% ethanolic extract of Abrusprecatorius were soaked respectively in 100mL distilled water and 70% ethanol. This mixture was decanted in a bulb of 500mL with 100mL of cyclohexane. The solution was shaked for 20mn then was decanted. The cyclohexane fraction was collected and evaporated. The aqueous superior phase was extracted with 100mL of dichloromethane. The dichloromethane fractions were collected and evaporated. The aqueous superior phase was extracted at last with 100 mL of ethyl acetate. The ethyl acetate fractions were collected and evaporated. The different fractions were tested on the growth of Salmonella typhimurium.

Evaluation of antisalmonella activity Determination of MIC

The minimal inhibitory concentration (MIC) was determined by broth dilution method. The minimal inhibitory concentration (MIC) was determined according Wilkinson and Gentry¹³. Two fold dilutions of the extract were made in the concentration range of 1, 25 mg/ml to 80mg/ml. The tubes were inoculated with a microorganism suspension at a final density of 10^6 cells/mL. The tubes were incubated at 37°C for 24h. The lowest concentration of the tube which did not show any visible growth after macroscopic evaluation was considered as the MIC.

RESEARCH ARTICLE



Ramanou AA et al, The Experiment, 2014., Vol. 27(1), 1844-1851

INTERNATIONAL JOURNAL OF SCIENCE AND TECHNOLOGY

Determination of MBC

The minimal bactericidal concentration (MBC) is defined as the concentration producing a 99.99% reduction in colony forming units (CFU) number in the initial inoculum. It was determined by subculture on nutrient agar as previously described¹⁴. The tubes without growth after 24 h of incubation were sub cultured on Mueller Hinton agar in Petri dishes for 24h.MBC was determined as the lowest concentration that showed no bacterial growth in the subcultures¹⁵.

Determination of antimicrobial activity

The antibacterial activity of aqueous, cyclohexane, dichloromethane, ethyl acetate and residual fractions of Thonningia sanguine and Abrusprecatorius against Salmonella typhimurium were performed by the agar disc diffusion method^{16,17}. The media (Mueller Hinton) along with the inoculum of Salmonella typhimurium(10^6 cfu/mL) was poured into the Petri plate. The disc (0,7 cm) was saturated with 50µL of the test compound, allowed to dry and then placed on the upper layer of the seeded agar plate. The plates were incubated at 37°C for 24h. Antibacterial activity was determined by measuring the inhibition zone diameters (mm) surrounding bacterial growth. The results were compared with the standard antibiotic: ciprofloxacin (5µg/mL).

All the antibacterial parameters have been determined after triplicate assays.

Statistical analysis

The data are presented as mean \pm SEM. All the data were analyzed by one-way ANOVA and differences between the means were assessed with Neuman-Keuls's multiple comparison tests. Differences were considered significant at p < 0, 05. All analyses were carried out using Graph Pad software, version 5.01 (USA).

RESULTSAND DISCUSSION

A total of 20 aqueous and 70% ethanol extracts from 10 different plant species were screened for antibacterial activity against two (2) clinical multi-drug resistant strains of Salmonella namely Salmonella typhi and Salmonella typhimurium. The results of the antisalmonella activity are shown in the table2.

The MIC values varied from 5 to 80 mg/mL and MBC values from 10 to 80 mg/mL. The aqueous extract of Thonningiasanguinea and 70% ethanol extract of Abrusprecatorius had the same and the best MIC and MBC values respectively5 mg/mL and 10 mg/mL against the two multi-drug resistant strains of salmonella. The aqueous extracts of Abrusprecatorius, Tectonagrandis,

Vernoniaamygdalina Nauclealatifolia and Olaxsubscorpioides presented MIC values of 40 mg/ml against Salmonella typhimurium. The 70% ethanol extracts of Nauclealatifolia and Ageratum conizoides exhibited the same MIC and MBC values respectively40 mg/ml and 80 mg/ml against Salmonella typhi, and Salmonella typhimurium.

The 70% ethanol extracts of Acanthosperiumhispidium and Manihotesculenta presented MIC values of 80 mg/ml against Salmonella typhi. The 70% ethanol extracts of Tectonagrandis exhibited MIC values of 80 mg/ml and MBC values of 80 mg/ml against Salmonellatyphimurium. The aqueous extract of Acanthosperiumhispidium presented MIC values of 40 mg/ml against Salmonellatyphimurium. The extracts of Cycasrevoluta, the aqueous extract of Ageratum conizoides, Manihotesculenta, the 70% ethanol extract of Vernoniaamygdalina showed no antibacterial activity against all the strains. The above results showed that eight (8) of the tested plants possessed weak antisalmonella activity against the multi-drug resistant tested strains. Two plants, namely Thonningiasanguinea and Abrusprecatoriusare the more potent antibacterial plants among the tested one against the two multi drug resistant MDR strains of salmonella according to the MIC and MBC values.

The aqueous extract of Thonningia. sanguinea has previously showed good antibacterial activities against extended spectrum-ß-lactamases (ESBL) strains of Klebsiella pneumonia and Escherichia coli with MIC of 6.25 mg/mL18. Moreover, N'guessan et al.10have also pointed out the antimicrobial activity of Thonningiasanguinea against Salmonella enterica (DT 104). Our results are in accordance

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RESEARCH ARTICLE



Ramanou AA et al, The Experiment, 2014., Vol. 27(1), 1844-1851

INTERNATIONAL JOURNAL OF SCIENCE AND TECHNOLOGY

with these results and provide a supplementary proof of the presence of compounds with promising antimicrobial activities against MDR strains of bacteria.

Furthermore, the work of Bolou et al.9 showed that these plants possessed interesting antibacterial activities on sensitive strains of Salmonella typhi and Salmonella typhimurium. Our results showed that these plants are also inhibitors of the growth of MDR strains of Salmonella.

The results on antibacterial activity of different fractions of Thonningiasanguinea and Abrusprecatorius were shown in table 3. The aqueous extract of Thonningiasanguinea showed the highest significant activity (11 ± 0.57) followed by the ethyl acetate fraction of Thonningiasanguinea ($10,3\pm0,4$), the ethyl acetate fraction of Abrusprecatorius (10 ± 00), the residual fraction of Thonningiasanguinea ($9\pm 1,15$), 70% ethanol extract of Abrusprecatorius ($9\pm1,0$), the residual fraction of Abrusprecatorius ($8\pm0,57$), the dichloromethane fraction of Abrusprecatorius ($7,6\pm0,33$) and the cyclohexane fraction of Thonningiasanguinea ($6,6\pm0,33$) against the MDR strain of SalmonellaTyphimurium. The cyclohexane fraction of Abrusprecatorius showed no pronounced antibacterial activity against Salmonella typhimurium.

The results showed no significant improving of the antisalmonella activities of the aqueous extract of Thonningiasanguinea and 70% ethanol extract of Abrusprecatorius.

This study may not be adequate to suggest potential antibioticagent considering the zone of inhibition which could be affected by the solubility and rate of diffusion in agar medium or its volatilization which could affect the results. However, this approach could be considered as preliminary step to find out promising candidates19. It is essential to apply other fractionation methods.

CONCLUSION

This study indicates that Thonningiasanguinea and Abrusprecatorius have the potential to generate novel antimicrobials metabolites against MDR strains of salmonella. Thonningiasanguinea and Abrusprecatorius can provide alternative solution for the treatment of salmonellosis particularly in Côte d'Ivoire.



RESEARCH ARTICLE

Ramanou AA et al, The Experiment, 2014., Vol. 27(1), 1844-1851

INTERNATIONAL JOURNAL OF SCIENCE AND TECHNOLOGY

species(family)	voucher number	Plant part tested	local name	popular use
Abrus precatorius Linn. (Fabaceae)	CNF/875	Seed	Alobogna	Dermatose, wound,cough
Acanthosperiumhispidium Schrank.(Asteraceae)	CNF/16762	Leaf	Lukoubassa moni	Paludism, typhoidfever
Ageratum conizoides Linn. (Asteraceae)	CNF/635	Leaf	Koitndré	Typhoidfever
Cycas revolutaThunb. (Cycadaceae)	CNF/1235	Leaf	efou	Typhoidfever
ManihotesculentaCrantz (Euphorbiaceae)	CNF/9024	Leaf	Agbagna	Paludism, Typhoidfever
Nauclea latifolia SM. (Rubiaceae)	CNF/15927	Leaf	Tôlé	Typhoidfever
OlaxsubscorpioidesOliv. (Olacaceae)	CNF/10756	Leaf	ifon	Paludism
Tectona grandis Linn. (verbenaceae)	CNF/842	Leaf	Tec	Typhoidfever
ThonningiasanguineaVahl. (Balanophoraceae)	CNF/17954	Flower	Glouglan	Typhoidfever
Vernonia amygdalina Del. (Asteraceae)	CNF/11694	Leaf	Abohoui	Paludism, Typhoidfever

References for uses: 20 and 21

Table 1: List of medicinal plants used in the antisalmonella assay



Ramanou AA et al, The Experiment, 2014., Vol. 27(1), 1844-1851

Table 2: Antibacterial activities of aqueous and 70% ethanol extracts of different medicinal plants against Salmonella strains.

				MIC mg/ml		MBC mg/ml	
Family	Name of Plant	Plant part	Extract	S.Typhi	S.Typhim	S.Typhi	S.Typhim
Balanophoraceae	Thonningia	F	AE	5	5	10	10
	sanguinea		ETH	80	80	80	80
Fabaceae	Abrus	S	AE	80	40	> 80	> 80
	precatorius		ETH	10	5	10	10
Verbenaceae	Tectona	L	AE	> 80	40	> 80	> 80
	grandis		ETH	> 80	80	> 80	80
Asteraceae	Vernonia	L	AE	> 80	40	> 80	> 80
	amygdalina		ETH	> 80	> 80	> 80	> 80
Olacaceae	Olax	L	AE	> 80	40	> 80	> 80
	subscorpioides		ETH	> 80	> 80	> 80	> 80
Rubiaceae	Nauclea	L	AE	80	40	80	> 80
	latifolia		ETH	40	40	40	40
Asteraceae	Acanthosperium	L	AE	> 80	40	> 80	> 80
	hispidium		ETH	80	> 80	80	> 80
Cycadaceae	Cycas	L	AE	> 80	> 80	> 80	> 80
	revoluta		ETH	> 80	> 80	> 80	> 80
Euphorbiaceae	Manihot	L	AE	> 80	> 80	> 80	> 80
	esculenta		ETH	80	> 80	80	> 80
Asteraceae	Ageratum	L	AE	> 80	> 80	> 80	> 80
	conizoides		ETH	80	80	80	80

AE= AqueousExtra	act	L= Leaves	MIC= Minimal Inhibitory Concentration
ETH= 70% Ethance	ol Extract	S= Seed	MBC= Minimal Bactericidal Concentration
		Typhim= Typhim	nurium F=
S.= Salmonella	Typ= Typhi	Flower	

RESEARCH ARTICLE



Ramanou AA et al, The Experiment, 2014., Vol. 27(1), 1844-1851

INTERNATIONAL JOURNAL OF SCIENCE AND TECHNOLOGY

Table 3: Antibacterial activities of differents fractions of Thonningia sanguine and Abrusprecatorius against S. Typhimurium.

Name of the plant	Fractions	Diameter of inhibition zone (mm) S.Typhimurium
	Aqueous	$11 \pm 0,57$
	Cyclohexane	6,6±0,33
Thonningiasanguinea(Fl)	Dichloromethane	8
	Ethyl acetate	10,3±0,4
	Residue	9± 1,15
	70% ethanol	9± 1,0
	Cyclohexane	-
Abrusprecatorius (S)	Dichloromethane	7,6±0,33
	Ethyl acetate	10
	Residue	8± 0,57
Ciprofloxacin 5µg		35

Fl: Flower, S: Seed, -: No Activity, S: Salmonella Note: Values are higly significant at 5% level

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RESEARCH ARTICLE



Ramanou AA et al, The Experiment, 2014., Vol. 27(1), 1844-1851

INTERNATIONAL JOURNAL OF SCIENCE AND TECHNOLOGY

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RAMANOU AA^{1*}, AHUA KM¹, YAPI HF¹, BOLOU GEK¹, DJAMANAJ^{1, 2}ANDN'GUESSAN JD¹

¹Biochemical Pharmacodynamy Laboratory, Department of Biosciences,

Felix HouphouetBoigny University,

Abidjan, Côte d'Ivoire

²Department of clinical and fundamental biochemistry, Pasteur Institute,

Abidjan,Côte d'Ivoire

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